

PRODUCT INFORMATION

Mouse anti-cMyc tag mAb sepharose Purification kit

v. 230201

For mammalian expression system

Catalog number C07001-K01 / C07001-K02

Package 5 rxns /10 rxns

Description

Affinity purified anti-cMyc antibodies, developed in mouse, were conjugated to NHS-sepharose. It is an efficient technique for isolating recombinant proteins or mammalian expression proteins. The cMyc epitope system relies on the peptide (N-Glu-Gln-Lys-Leu-Ile-Ser-Glu-Glu-Asp-Leu-C), which is able to react with N- and C- terminal cMyc tagged fusion proteins and may be used for the immunoprecipitation or immune affinity purification.

The purified antibody is immobilized at 3-5 mg antibody per mL of 50% slurry and this kit allows a rapid and efficient affinity purification of active cMyc fusion proteins. The affinity resin allows an efficient binding of cMyc fusion proteins without the need for preliminary steps and calibrations. The affinity bound cMyc fusion proteins can be efficiently eluted from the resin by acid condition. The eluted proteins can be used for characteristic analysis.

Component	Reagents & Materials	Quantity for 10 rxns (C07001-K02)	Quantity for 5 rxns (C07001-K01)	composition
		Mouse anti-cMyc tag mAb sepharose	2 mL X 1 vial	1 mL X 1 vial
	Wash Buffer (10X concentration)	5.0 mL X 1 vial	2.5 mL X 1 vial	100 mM Tris/Cl pH 7.5; 1.5 M NaCl; 5 mM EDTA
	Elution Buffer	10 mL X 1 vial	5 mL X 1 vial	0.1 M Glycine pH 2.7
	Neutralization Buffer	2 mL X 1 vial	1 mL X 1 vial	2 M Tris pH 8.0
	spin column	10 pcs	5 pcs	
	collection tube	10 tubes	5 tubes	

Note: Sepharose is 1:1 suspension in Phosphate Buffered Saline, pH 7.4, containing 0.05% sodium azide as a preservative.

Product capacity

The binding and elution capacity of 1 mL Mouse anti-cMyc tag mAb sepharose are commonly more than 1 mg of cMyc fusion protein. Trying different elution buffers for optimal results is recommended.

**Materials Required
but Not Provided**

- Micro-centrifuge capable of 15,000 x g
- 1.5 mL Centrifuge tubes
- End-over-end rotator
- ColP Lysis Buffer (mild reaction): 10 mM Tris/Cl pH 7.5; 150 mM NaCl; 0.5 mM EDTA; 0.5% NP-40
- RIPA (vigorous reaction): 100 mM Tris/Cl pH 7.5; 300 mM NaCl; 0.2% Sodium Deoxycholate (or 0.1% SDS); 2% NP-40

Storage

For sustainable use and long term storage, store at 2 °C to 8 °C. **DO NOT FREEZE.**

Precautions**Disclaimer**

This product is for R&D use only, not for drug, household, or other uses.

General notes

The Mouse anti-cMyc tag mAb sepharose is stored in Phosphate Buffered Saline containing 0.05% sodium azide. The PBS must be removed before use and the resin should be equilibrated with 1X Wash Buffer. The equilibration can be performed at room temperature or at 2-8 °C. The Wash Buffer is original stock concentration. Dilute Wash Buffer (10X concentration) to 1X working concentration with distilled water.

In the case of bulk reaction. Users can make a pre-reaction through mixing sample and sepharose in 15 mL / 50 mL tube, and then transfer the mixture into the column for binding.

ProcedureSuggestions on purification of cMyc fusion proteins

1. Cellular debris and particulate matter must be removed by centrifugation or filtration prior to purification on the column.
2. Highly viscous samples which may contain chromosomal DNA or RNA should be sonicated or treated with nuclease to decrease the viscosity.
3. Perform all steps on ice.

A. Sample preparation (Lysis of Mammalian Cells)

1. Detach the cells from the culture dish and collect the cell suspension into the centrifuge tube.
2. Centrifuge the cell suspension at 400 x g for 5 minutes to pellet the cells. Carefully remove and discard the supernatant.
3. Wash cells by re-suspending the cell pellet in ice-cold PBS.

4. Centrifuge the cell suspension at 400 x g for 5 minutes to pellet the cells. Carefully remove and discard the supernatant.
5. Add 200 μ L of Lysis buffer to the cell pellet and vortex.
6. Incubate the sample for 15 minutes on ice.
7. Remove cell debris by centrifugation at 15,000 x g for 5 minutes at 4°C.

B. Column preparation

1. Place an empty spin column on the collection tube.
2. Wash the column with 200 μ L Wash Buffer.
3. Allow the buffer to drain from the column and leave residual Wash PBS in the column to aid in packing the Mouse anti-cMyc tag mAb sepharose, then discard the buffer in the collection tube.

C. Packing the Column

1. Completely suspend the vial of Mouse anti-cMyc tag mAb sepharose.
2. Transfer 200 μ L volume to an empty centrifuge tube, and wash the sepharose with 1 mL Wash Buffer.
3. Spin down the sepharose with 100 x g, 30 seconds' centrifugation and discard supernatant.
4. Immediately transfer the sepharose to the spin column. Allow the sepharose bed to settle. Please prevent the sepharose bed from getting dried.
(Note: Make sure the column filter is fixed in the correct position before transferring the sepharose).

Procedure

D. Binding cMyc fusion protein to the column

1. Dilute the sample with Wash Buffer in 1:3 proportion.
2. Load the sample on the spin column and centrifuge the column at 100 x g for 30 seconds. Users can also perform this binding reaction in a new 1.5 mL centrifuge tube.
(Note: Depending upon the cMyc fusion protein and the flow rate, not all of the protein may bind. Repeat loading the sample to increase binding efficiency).
3. Collect the fractions using empty centrifuge tube.
4. Wash the spin column with 300 μ L Wash Buffer more than 6 times.
(Note: To eliminate the noisy band in sample, more washing step is recommended).

E. Elution of cMyc fusion protein

1. Add 5 x 100 μ L Elution Buffer to elute the bound cMyc fusion protein from the spin column to the collection tube. This step can be supported by a centrifugation at 100 x g for 30 seconds.
2. Immediately neutralize the eluted sample by adding 10 μ L Neutralization Buffer. Assay sample concentration by measuring the absorbance at 280 nm and combine the fractions with highest absorbance.
(Note: Measuring the absorbance after each Elution move can help collecting the sample more accurately).

F. Optional instead of elution step

Resuspended Mouse anti-cMyc tag mAb sepharose in 100 μ L 2 x SDS-Sample Buffer for 10 minutes at 95°C to dissociate immune-complexes from Mouse anti-cMyc tag mAb sepharose. Mouse anti-cMyc tag mAb sepharose can be collected by centrifugation at 2500 x g for 2 minutes at 4°C and SDS-PAGE is performed with the supernatant.

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